

An Elimination–Rearrangement of Ribulose-1,5-bisphosphate with Implications for Riboflavin Biosynthesis

David H. G. Crout and John A. Hadfield

Department of Chemistry, University of Warwick, Coventry CV4 7AL, U.K.

Ribulose-1,5-bisphosphate (5) undergoes a sequence of elimination and benzilic acid rearrangement in a process with features that parallel those of the rearrangement occurring during incorporation of pentose precursors into riboflavin (1).

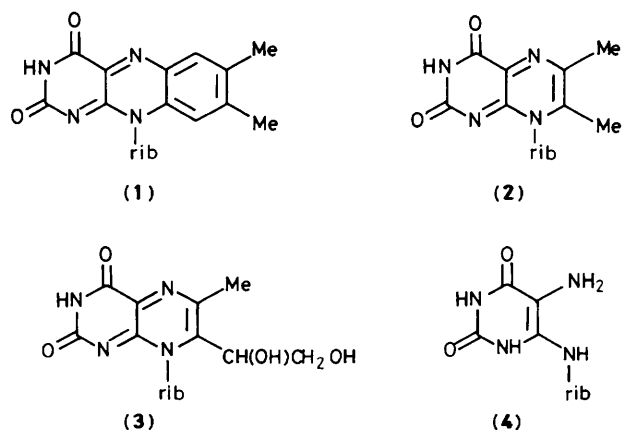
The biogenesis of the *o*-xylene moiety of riboflavin (1) has not been firmly established. Although the later steps from 6,7-dimethyl-8-ribityl-lumazine (DMRL) (2) are well understood,¹ the origin of the four-carbon component in DMRL (2) which furnishes all eight carbon atoms of the *o*-xylene system of riboflavin (1) is still uncertain. Previously postulated four-carbon precursors such as acetoin (3-hydroxybutan-2-one)² and diacetyl (butane-2,3-dione)³ have now been eliminated although rumours of their involvement still persist.⁴

Bresler *et al.* observed that a riboflavin-requiring mutant of *Bacillus subtilis* excreted a compound identified as 6-methyl-7-(1,2-dihydroxyethyl)-8-ribityl-lumazine (MERL) (3) which supported the growth of a riboflavin auxotroph, albeit at a concentration 10³ times greater than that required when riboflavin itself was supplied.⁵ It was suggested that the C₅ unit of this compound was derived from the ribityl component of the established riboflavin precursor 5-amino-2,4-dioxo-6-ribitylamino-pyrimidine (ADRAP) (4). Mechanistically and

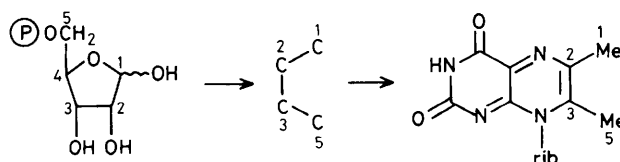
biochemically acceptable processes can be postulated whereby MERL (3) might be converted into DMRL (2) and thence into riboflavin (1) with loss of the hydroxymethyl group. Although the evidence supporting the precursor role of MERL (3) in riboflavin (1) biosynthesis seemed persuasive, it was not consistent with the results of Floss *et al.* who have obtained evidence that a pentose precursor or a closely related species, is converted into the C₄ unit of DMRL (2) with excision of C-4 and formation of a new carbon–carbon bond between C-3 and C-5 (Scheme 1).⁶

We here describe experimental evidence bearing both on MERL (3) formation and on the rearrangement mechanism of riboflavin (1) biosynthesis proposed by Floss *et al.*⁶ These experiments arose out of mechanistic considerations pointing to the possible involvement of ribulose-1,5-bisphosphate (5), or an equivalent system, in riboflavin (1) biosynthesis. Thus in Scheme 2, a postulated sequence is shown whereby ribulose-1,5-bisphosphate (5) might be converted into the 5-phosphate of the diketone (6). Hydrolysis of the 5-phosphate ester followed by condensation with ADRAP (4) would furnish MERL (3). This sequence requires Lobry de Bruyn–Alberda van Ekenstein rearrangement of ribulose-1,5-bisphosphate (5), phosphate elimination, for which there is ample chemical^{6,7} and biochemical⁸ precedent, and ketonisation of the resulting enol.

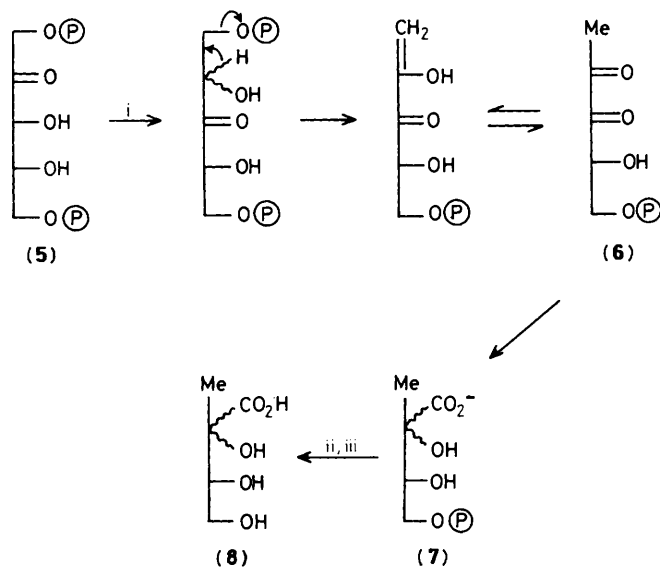
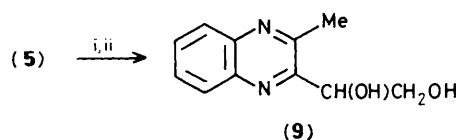
To test the validity of this proposal ribulose-1,5-bisphosphate (5) was incubated at pH 13 and product formation was monitored by ¹³C and ¹H n.m.r. spectroscopy.



rib = ribityl



Scheme 1

Scheme 2. Reagents: i, pH 13; ii, alkaline phosphatase; iii, H⁺

Scheme 3. Reagents: i, pH 6.2, 1,2-diaminobenzene; ii, alkaline phosphatase.

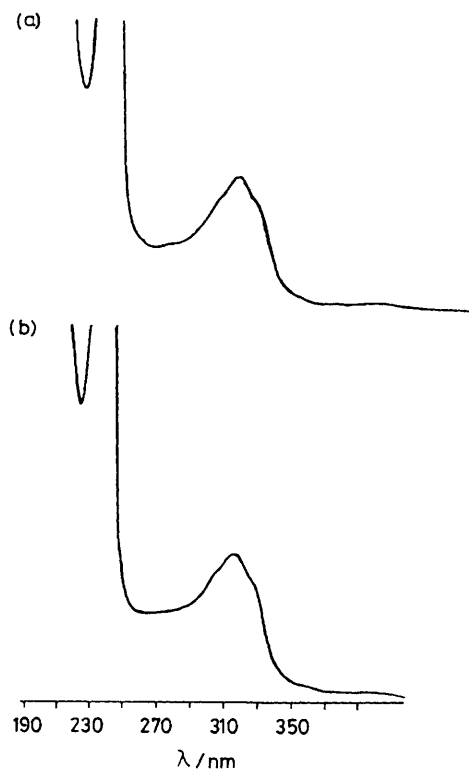
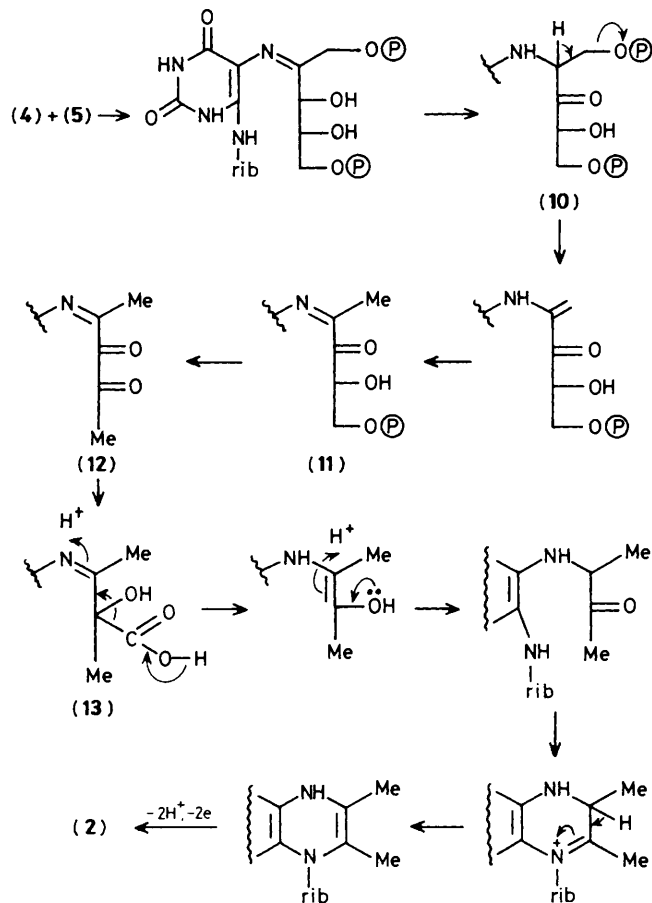


Figure 1. U.v. spectra [MeOH:H₂O-0.05% CF₃CO₂H (1:1)] of 2-(1,2-dihydroxyethyl)-3-methylquinoxaline (9) (a) from the incubation of ribulose-1,5-bisphosphate with 1,2-diaminobenzene at pH 6.2; (b) synthetic.



Scheme 4. rib = ribityl.

Signals attributable to newly formed methyl groups were observed but at higher field than expected for the diketone (6). Also, a carbonyl carbon resonance was observed in the ¹³C n.m.r. spectrum with a chemical shift (δ 181.7) corresponding to that of a carboxylate rather than a ketonic carbonyl group. It appeared likely that formation of benzylic acid rearrangement (6) was rapidly followed by benzilic acid rearrangement to the carboxylate (7). Proof was obtained by hydrolysis to the trihydroxy acid (8) using alkaline phosphatase. This acid was obtained as a mixture of diastereoisomeric lactones identified by comparison with an authentic sample. Comparison was made by ¹H n.m.r. and by g.l.c.-mass spectral analysis† of trimethylsilyl derivatives of both the lactone and the corresponding hydroxy acid.

When the incubation of ribulose-1,5-bisphosphate (5) was carried out at pH 6.2 in the presence of 1,2-diaminobenzene a complex mixture resulted which was subjected to hydrolysis with alkaline phosphatase. From the product mixture a compound was isolated by h.p.l.c. which had a retention time identical (confirmed by co-chromatography) with an independently synthesised sample of the MERL (3) analogue (9) (Scheme 3) and with a qualitatively identical u.v. spectrum (Figure 1). It therefore appears that under these conditions, the intermediate (6) postulated to have been formed from ribulose-1,5-bisphosphate (5) can be trapped as the quinoxaline (9). Alternatively, the elimination may occur in the Schiff

† Determined using a Kratos MS 80 mass spectrometer with g.l.c. on SE 30, He carrier gas, temperature programme 120°/3° min⁻¹/180°C.

base formed by initial condensation between ribulose-1,5-bisphosphate (**5**) and 1,2-diaminobenzene.

Floss *et al.*⁶ have proposed a radical mechanism for the rearrangement (Scheme 1) of the postulated five-carbon precursor of the *o*-xylene system in riboflavin (**1**). However, the above results suggest an alternative mechanism. Thus condensation of ribulose-1,5-bisphosphate (**5**) with ADRAP (**4**) (Scheme 4) followed by Amadori rearrangement would give the iminoketone (**10**). Phosphate elimination would then lead to the iminoketone (**11**) which has the functionality that permitted the elimination-benzilic acid sequence to occur with ribulose-1,5-bisphosphate (**5**) itself. However, in this case a β -iminocarboxylic acid (**13**) would be produced which could undergo ready decarboxylation. Formation of DMRL (**2**) would then require only condensation and two-electron oxidation. To be consistent with the results of Floss *et al.*,⁶ it is only necessary to make the further postulate of enzymatic control of methyl group rather than enamino group migration during the benzilic acid rearrangement (**12**) \rightarrow (**13**).

The sequence of Scheme 4 is open to considerable modification with regard to detail. However, its basic features, generation of a 1,2-diketone by elimination in a 3-oxophosphate, benzilic acid rearrangement, and decarboxylation, offers an attractive explanation, consistent with the available evidence, for the conversion of a pentose precursor into the *o*-xylene ring of riboflavin (**1**).

Ribulose-1,5-bisphosphate (**5**) is a key intermediate, as the substrate for the enzyme ribulose-1,5-bisphosphate carboxylase, in the photosynthetic carbon cycle. The degradative rearrangement demonstrated above therefore has implications for ribulose-1,5-bisphosphate (**5**) metabolism,^{9,10} since phosphate elimination, as in our studies, can occur over a wide

pH range,¹¹ encompassing the region normally regarded as 'physiological.'

We thank the S.E.R.C. for financial support.

Received, 28th November 1986; Com. 1688

References

- 1 G. M. Brown and J. M. Williamson, *Adv. Enzymol.*, 1982, **53**, 345.
- 2 B. Bews and D. H. G. Crout, *J. Chem. Soc., Perkin Trans. 1*, 1986, 1459.
- 3 G. W. E. Plant, *Compr. Biochem.*, 1971, **21**, 11.
- 4 K. Nakajima, *Acta Vitaminol. Enzymol.*, 1985, **7**, 25.
- 5 S. E. Bresler, D. A. Perumov, T. P. Chernik, and E. A. Glazunov, *Genetika*, 1976, **12**, 83.
- 6 P. Nielsen, G. Neuberger, I. Fujii, D. H. Brown, P. J. Keller, H. G. Floss, and A. Bacher, *J. Biol. Chem.*, 1986, **261**, 3661.
- 7 T. C. Bruice, and S. Benkovic, 'Bioorganic Mechanisms,' vol. 2, W. A. Benjamin, Inc., New York and Amsterdam, 1966, pp. 25–29.
- 8 M. Flavin and T. Kono, *J. Biol. Chem.*, 1960, **235**, 1109; P. R. Srinivasan, J. Rothschild, and D. B. Sprinson, *ibid.*, 1963, **238**, 3176.
- 9 C. Paech, J. Pierce, S. D. McCurry, and N. E. Tolbert, *Biochem. Biophys. Res. Commun.*, 1978, **83**, 1084. (The authors proposed a sequence similar to that of Scheme 2 to explain the formation of an inhibitor of ribulose-1,5-bisphosphate carboxylase from xylulose-1,5-bisphosphate.)
- 10 A. Jaworoski, F. C. Hartman, and I. A. Rose, *J. Biol. Chem.*, 1984, **259**, 6783.
- 11 T. C. Bruice and S. Benkovic 'Bioorganic Mechanisms,' vol. 2, W. A. Benjamin, Inc., New York and Amsterdam, 1966, pp. 26–27.